East African Scholars Journal of Agriculture and Life Sciences

Abbreviated Key Title: East African Scholars J Agri Life Sci ISSN 2617-4472 (Print) | ISSN 2617-7277 (Online) Published By East African Scholars Publisher, Kenya

Volume-7 | Issue-5 | May-2024 |

Original Research Article

DOI:10.36349/easjals.2024.v07i05.001

OPEN ACCESS

Histological Study on Ovarian Development of Freshwater Prawn Macrobrachium dayanum (Henderson)

Krishan Raj Kant¹, Chanchal Angral², Wahied Khawar Balwan^{3*}

¹School Education Department, Jammu and Kashmir, India

²Department of Zoology, Govt. Degree College R. S. Pura, Jammu and Kashmir, India

³Department of Zoology, Govt. Degree College Doda, Jammu and Kashmir, India

Article History Received: 09.04.2024 Accepted: 11.05.2024 Published: 14.05.2024

Journal homepage: http://www.easpublisher.com



Abstract: The development of the oocyte into mature ova has been studied under two principal categories namely pre-vitellogenic and vitellogenic phase. Other than oocytes, oogonia have also been seen in the ovaries during different months of the year. It has been observed that the ovaries of *Macrobrachium dayanum* possess maximum percentage of oogonia during the months of December to January and June to July which indicate that they are passing through immature stage. It was further observed that during the month of April and October to November ovaries were filled with primary, secondary and tertiary vitellogenic oocytes.

Keywords: Oogonia, vitellogenic oocytes, ovaries, Macrobrachium dayanum.

Copyright © 2024 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

INTRODUCTION

Prawn fishery of India with an annual catch of over 100000 tons is second only to the United States and accounts for 18% of the world's population. Flourishing trades of exporting prawn pulp to Burma and Malaya from earlier times and frozen and canned prawns to USA and Japan has made Indian prawns a major foreign exchange earner (Jhingran 1991). The order decapoda contain the familiar shrimps, crayfish, lobsters, crabs and some of the most highly specialised crustaceans, the 8500 described decapods represent almost one third of the known species of crustaceans and they constitute the most important fishery of India. Most of the decapods are marine, but the crayfish, some shrimps and a number of crabs are the one that have invaded freshwater (Barnes, 1974). Holthius in 1980 listed 48 species of the genus Macrobrachium that are consumed as food and hence are commercially important. About half of these species have been examined for their potential suitability for aquaculture (Holthius 1980; New 1990, 1995; Brown 1991; Jhingran 1991; Jayachandran and Joseph 1992). Apart from being a delicacy they are a rich source of protein and vitamin (A and D) most needed to combat malnutrition in human population of the world. They contain considerable quantities of glycogen and free amino acids in their muscles, imparting their flesh a

sweet taste. As they contain a very little fat, they have become a favourite protein food for the weight conscious aristocrats (Kumar, 2003).

Of the three freshwater species, *Macrobrachium dayanum* qualifies the criteria of edibility on the basis of its size and habitat from freshwater of Jammu. Except preliminary studies conducted on its morphology, nutritional status and reproductive cycle (Kailoo 1984, Malik 2004, Chalotra 2004, Samyal 2006, Samyal *et al.*, 2007 and Manhas 2012) [12, 21, 20] nothing is on record with regard to reproductive biology of *Macrobrachium dayanum*. Realizing this, therefore, presently studies on maturation and ovarian development were undertaken.

MATERIAL AND METHODS

Experimental organisms Prawn, *Macrobrachium dayanum* were collected from their natural habitat from a stream at Gho-Manhasan situated at 32° 67' Lat N; 74° 79' Long E located at a distance of 20 km North West of Jammu city. Monthly collections were made with the help of rectangular haul / sweep net with 1620 cm sq. mouth area (1mm mesh size) and 80 cm long during morning hours (8:00-12:00). Most of the live specimens were collected from Gho-Manhasan stream because of easy access and availability in abundance throughout the year. On the basis of sexual dimorphic characters, sex wise segregation was done in case of Macrobrachium dayanum. Fresh specimens of prawns were dried by wrapping in blotting paper and weighed (in grams) using electronic weight balance. Five to six specimen of prawn were sacrificed and their gonads were removed and weighed on electronic balance. Gonads (ovary) were fixed in freshly prepared Bouine's solution (Picric acid 70%, Acetic acid 25% and formalin 5%) for 12-24hrs. After fixation, gonads were washed 3-4 times with tap water, then processed through routine alcoholic grades from 70% (5-10minutes), 90% (5-10minutes), 100% (5-10 minutes) and finally cleared in xylene. The material was then transferred in xylol wax and kept overnight at 40°c for effective impregnation of the wax. Blocks of gonadal tissue were then prepared for microtomy. 5-8µ paraffin sections were stained using Mallory's triple & Haemotoxyline-eosine. Finally the material was mounted with DPX and coverslips. Slides so prepared were then studied under Nikon Ys 100 microscope and photographed with the help of SDC-313-Camera.

RESULTS AND DISCUSSION

The development of the oocytes into the mature vitellogenic ova have been studied under following two principal categories. Other than oocytes, oogonia have also been seen in ovaries during different months of year.

- 1. Pre-vitellogenic phase.
- 2. Vitellogenic phase.

A brief description of these stages is as follows;

Oogonia

These are small spherical cells with centripetally located nucleus. Oogonia possess homogenous cytoplasm. These have been observed to be present either near the germinal zone attached to the germinal epithelium or freely lie in the ovocoel in the months from December to March and May to July. Percentile occurrence of these oogonia varied from 59.13% to 63.18% during December to January and 54.17% to 62.03% during months of June to July (Table 1) (Figure 1). These oogonia start declining from the month of February and August onwards to touch as low as 4.73% in the month of May and 5.13% in the month of November (Table 1).

Stage I or Previtellogenic oocytes

The cytoplasm of these oocytes have been observed to be homogenous as all the oocytes lacked yolk. The nucleus appeared vesicular and nuclear material stained black with haemotoxyline stain. Previtellogenic oocytes are comparatively larger in size and lie free in the ovocoel (Figure 2). The oocytes which range in diameter from 0.140- 0.189 mm showed great variation in size. Based on size variation, these have been categorized into previtellogenic oocytes-I (PVOI) (0.140-0.156 mm), pre- vitellogenic oocytes-II (PVOII) (0.158-0.172 mm) and pre- vitellogenic oocytes-III (PVOIII) (0.172-0.189 mm) with a mean oocyte diameter of 0.152 mm, 0.166 mm and 0.184 mm respectively (Table 2 and Figure 2). Percental occurrence of all three types of pre-vitellogenic oocytes have been observed to vary from 33.12- 38.19% during the months of December to January and 37.18-43.14% during the months of June to July with maximum percental occurrence of 43.14% in the month of June. These oocytes start declining in the ovaries from the months of January and June onwards when their contribution is just 4.17% in the month of October and 3.27% in the month of April (Table 2).

Vitellogenic Oocytes

On the basis of size of oocytes, nuclear changes, appearance of yolk vesicles and yolk accumulation, vitellogenic oocytes have been categorized into three sub-stages. (I) Primary vitellogenic oocytes (S-II), (II) Secondary vitellogenic oocytes (S-III) and (III) Tertiary vitellogenic oocytes (S-IV) / mature oocytes. This is the phase when synthesis of yolk takes place. Follicle cells start appearing during this phase and marked changes in the nucleus, nucleolus and ooplasm have been observed.

Stage II or Primary Vitellogenic Oocyte

Ooplasm of these oocytes is granular, small yolk vacuoles have been observed to make their appearance in the periphery of ooplasm of the oocytes. Size of the oocytes as observed ranges in diameter from 0.196-0.214 mm. Nucleus stain bluish black with haemotoxyline, is centrally located, increases in size and had a wavy nuclear membrane. During this stage percental occurrence of these small size primary vitellogenic oocytes have been observed to vary from 34.16 - 41.12% & 36.21- 47.38% during the months of February to March and August to September respectively (Table 2). Small round follicular cells around oocytes have been observed.

Stage III or Secondary vitellogenic oocytes

In secondary vitellogenic oocytes, small amount of yolk get deposited in vacuoles, small unstainable vacuoles fuse together to form large yolk vesicles. Small eosinophilic yolk granules start accumulating in the periphery region of oocortex. The follicle cells remain confined to the periphery as was characteristic in the earlier phase. With advancement in this phase of vitellogenic oocytes, the perinuclear ring disappears. Nucleus in these oocytes increases further in size and nucleoli start appearing in the nucleoplasm. In some oocytes towards the end of this stage the nuclear membrane become indistinct. The diameter of secondary vitellogenic oocytes vary from 0.198- 0.242mm. Their occurrence in the ovaries have been recorded to be in the range of 49.05% - 54.03% and 48.31- 57.13% during the months of April to May as well as during the months of September to October respectively (Table 2) (Figure 3).

Stage IV or Tertiary vitellogenic oocyte / Mature oocytes

Tertiary vitellogenic oocytes further increase in size and attain their utmost size of 0.198 - 0.264 mm during this phase. Yolk vesicles are distributed in the entire cytoplasm and nucleus is displaced towards the periphery which acquires eccentric position. Yolk deposition is almost complete in all these oocytes and whole of the ooplasm upto perinuclear region gets occupied by yolk globules. These oocytes are now designated as mature ova. The entire cytoplasm becomes eosinophilic. Percental occurrence of tertiary vitellogenic oocyte vary from 42.86 % - 64.15 % & 43.16 - 61.73% in the months of May as well as during the months of November to December (Table 2) (Figure 4).

Fully mature oocytes called, mature ova, ovulate and get deposited in the brood pouch of the ovigerous prawns in the 1st, 2nd & 3rd pleopods where the ova not only get fertilized but start further development also. The maximum preponderance of mature stage IV oocytes during the months of May to June as well as November to December signifies that prawn *Macrobrachium dayanum* is a biannual breeder.

The present viewpoint find support from the findings of Subaramanium (1963), Ryan (1965), Jegla (1966), Kulkarni (1992), Kailoo (1984), Malik (2006), Samyal et al., (2006) [21] and Samyal (2007). Concerning different stages during development of oocytes into ovum, present author's view point get strengthened from the observations recorded by a number of authors on Macrobrachium dayanum (Adiyodi and Adiyodi, 1974; Subaramanium, 1979; Kailoo, 1984; Malik, 2006; Samyal et al., 2006 and Samyal, 2007) as well as those who have worked on different species (Joshi, 1980 on Peaneus stylifera and hardwickii; Mirajaker, 1980 Penaeus on kistensis; Garimella, 1982 Macrobrachium on Macrobrachium lamarrei) and Chakarvarty (2003) who too reported similar stages in the development of oocytes into ovaries of giant freshwater prawn Macrobrachium rosenbergii. Observations made by these workers are in line with present observations too wherein they have recorded four different stages in the ovarian development cycle of respective prawn species.

 Table 1: Maturity stages, percental occurrence, oocyte diameter colour of ovary and macroscopic characteristics of ovary of Macrobrachium dayanum

Maturity Stage /	Most prevalent	Oocyte	Colour of	Macroscopic characteristics of ovary
Month	oocytes	Diameter	ovary	-
	development stage		-	
	with its percental			
	occurrence			
Immature	Oo = 59.13 -	SI = 0.140	Translucent	Ovary is small, translucent and difficult
undeveloped phase	63.81%.	- 0.189		to distinguish. Oogonia cells are
/ oogonial stage	Oo = 54.17 - 62.03%	mm.		aggregated, as seen through the
(Dec-Jan),	S-I = 37.18 - 43.14%	SII = 0.196		microscope.
(June-July)	S-II = 33.12 –	- 0.214 mm		
	38.19%			
Previtellogenic	SIII = 7.05 - 9.95%.		Yellowish	Oogonia cells have advanced into
phase/	Rest were SI & SII.	SIII =		oocyte stage. Developing ovaries are
Developing phase		0.198-		larger, opaque, and yellowish, with
(Feb-Mar),		0.242 mm.		scattered melanophores over the
(Aug-Sep)				surface. Pre-vitellogenic oocytes
				appear. Ovaries are yellow is in colour.
Vitellogenic phase/	SIII = 49.05-	SIV =	Dark orange	Vitellogenesis shown by eosin stained
Developed phase.	54.03%	0.198-	C C	oocytes. Nucleus appears. Vesicular
(April), (Oct-Nov)	SIV = 38.11-	0.264 mm.		bodies appear with chromatin clumps.
	45.35%. Rest were			Ova are nearly ripe. Ovaries are dark
	SI & SII.			orange in colour.
Mature phase.	SIII = 38.05-	SIV =	Dark green	Post vitellogenic stage is characterised
(May-June), (Dec).	47.34%	0.242 -	Ŭ	by shrunken and irregularly shaped
	SIV = 42.86-64.15%	0.264 mm.		cells observed after ova release. Ovary
	Rest were SI & SII.			is dark green in colour.
	Oo = 4.73% &			Ŭ
	5.13%			

Oo = oogonia, S-I = Previtellogenic oocytes, S-II = Primary vitellogenic oocyte, S-III = Secondary vitellogenic oocyte & S-IV = Tertiary vitellogenic oocyte

Stages	Oocyte	Oocytes			Months
	diameter	PVO I	PVO II	PVO III	
		0.140 mm -	0.158 mm-	0.174 mm-	
		0.156 mm	0.172 mm	0.189 mm	
Pre-vitellogenic	Percental	33.12 - 38.19			December – January
Stage (S-I)	occurrence	37.18 - 43.14			June – July
		4.17			October
		3.27			April
Primary vitellogenic	Oocyte diameter	0.196 mm - 0.214 mm 34.16 - 41.12			
Stage (S-II)	Percental				February – March
	occurrence	36.21 - 47.38			August – September
Secondary vitellogenic	Oocyte diameter	0.198 mm - 0.24	2 mm		
Stage (S-III)	Percental	49.05 - 54.03			April – May
	occurrence	48.31 - 57.13			September – October
Tertiary vitellogenic	Oocyte diameter	0.198 mm - 0.26	64 mm		
Stage (S-IV)	Percental	42.86 - 64.15			May
	occurrence	43.16 - 61.73			November - December

Table 2: Maximum percental occurrence and oocyte diameter of previtellogenic and vitellogenic oocytes observed
during the year

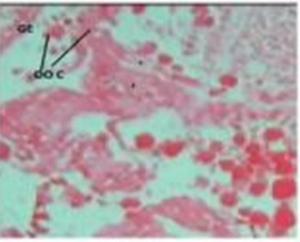


Fig 1: Microphotograph of T.S of ovary showing oogonia attached to germinal epithelium in the ovocoel

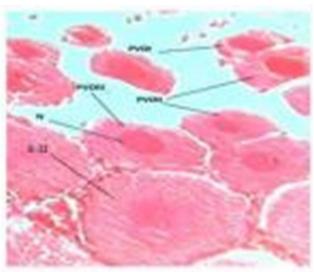


Fig 2: Microphotograph of T.S of ovary showing three types of previtellogenic oocytes (PVOI = Pre-vitellogenic oocyte sub-stage – I, PVOII = Pre- vitellogenic oocyte sub-stage – II, PVOIII = Pre-vitellogenic oocyte sub-stage – III, N = Nucleus, III = Stage – III vitellogenic oocyte)

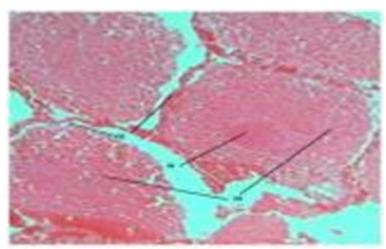


Fig 3: Microphotograph of T.S of ovary showing primary vitellogenic oocytes (FE = Follicular epithelium, N = Nucleus, II = Stage – II vitellogenic oocyte)

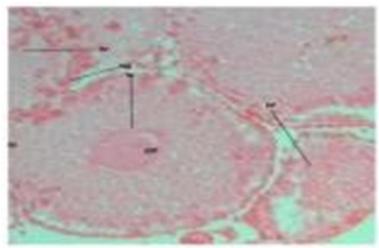


Fig 4: Microphotograph of T S of ovary showing secondary & tertiary vitellogenic oocytes (III = Stage - III vitellogenic oocyte, IV = Stage-IV vitellogenic oocyte, N = Nucleus, Y G = Yolk globules C P = Cytoplasm)

REFERENCES

- Adiyodi, K. G., & Adiyodi R. G. (1974). Comparative physiology of reproduction in Arthropods: Comparative physiology and biochemistry. Edited by O. Lowenstein Academic Press, New York, London, 5, 37-107.
- Barnes, R. D. (1974). Invertebrate Zoology V.B. Saunders Company, 1-841.
- Brown, M. L. (1991). Freshwater Power. In production of aquatic animals, world animal science, CY, (ed. By C E Nash). Elsevier scientific publishing; Amsterdam, 31-43.
- Chakravorty, M. S. (2003). Studies on the oocyte stages of the prawn *Macrobrachium rosenbergii* de Man. *J Aqua Biol*, *18*(2), 101-106.
- Chalotra, R. (2004). Studies on early life history of freshwater prawn *Macrobrachium dayanum*. M. Phill. Dissertation Submitted to University of Jammu, Jammu, 2004.
- Dutta, S. P. S. (1978). Limnology of Gadhigarh stream (Miran sahib, Jammu) with special reference

to producers inhabiting the stream. Ph.D Thesis University of Jammu, Jammu.

- Garimella, G. R. (1982). Reproductive biology of freshwater prawn *Macrobrachium lamarrei*. Ph.D, Thesis, Marathawda Univ. Aurangabad.
- Hotlhius, L. B. (1980). Shrimps and prawn of the world and annotated catalogues of species of interest of fishers. FAO fish synopsis, 125, 1-261.
- Jayachandran, K. V., & Joseph, N. I. (1992). A key for the commercially important *Macrobrachium spp*. of India with a review of their bionomics in fresh water prawns. Proceedings of the national symposium on freshwater prawns (*Macrobrachium spp.*), 12-14. December, 1990, Kochi, (Ed, By E. G. Sciens), Kerala agricultural university thissur, 72-77.
- Jegla, T. C. (1966). Reproductive and molting cycles in cave crayfish. *Biol Bull*, *130*(3), 345-358.
- Jhingran, V. G. (1991). Fish and fishers of India, 3rdedn, Hindustan publishing Corporation (India); Delhi.

- Joshi, P. (1980). Reproductive physiology and Neurosecretion in some Indian marine prawn. Ph. D thesis, Marathwada University, Aurangabad.
- Jyoti, M. K., & Kailoo, U. C. (1985). Spawning Season of *Macrobrachium dayanum* (Henderson) inhabiting Jammu water, India. *Zoological Oriental's Z*, 45-48.
- Kailoo, U. C. (1984). Studies on systematics and reproductive biology of fresh water prawns of Jammu waters. M. Phill. Thesis, Jammu University, 1984.
- Kulkarni, M. Y. (1992). Studies on the biology of the freshwater prawn *Macrobrachium kistensis*. Ph.D Thesis, Marathwada University, Aurangabad.
- Kumar, H. D. (2003). Sustainability and management of agriculture and fisheries. Days publishing house New Delhi, 1-325.
- Malik, N. (2006). Studies on female reproductive system of *Macrobrachium dayanum* (Henderson).
 M. FSc. Dissertation Submitted to University of Jammu.
- Malik, N. (2006). Studies on female reproductive system of *Macrobrachium dayanum* (Henderson).
 M. FSc. Dissertation Submitted to Directorate of Distance Education, University of Jammu, 2006.
- Mirajkar, M. S. (1980). Studies on reproductive biology and neurosecretions of fresh water prawn

Macrobrachium kistensis. Ph.D Thesis, Marathawda University, Aurangabad.

- New, M. B. (1990). Freshwater prawn culture. A review. *Aquaculture*, 88, 99-141.
- New, M. B. (1995). Status of freshwater farming: A review. *Aquaculture research*, 26, 1-54.
- Ryan, E. P. (1965). A study of the reproductive biology of the hoale crab, *Portunus sanguinoleutus* (Herbst) Ph.D Thesis, Hawai University.
- Samyal, A., Bakhtiyar, Y., Verma, A., & Langer S. (2006). Studies on seasonal variations in lipid composition of muscles, hepatopancreas and ovary of freshwater prawn *Macrobrachium dayanum* (Henderson) during reproductive cycles. *Advanced Journal of Food Science and Technology*, 3(3), 160-164.
- Samyal, A. (2007). Seasonal dynamics in biochemical composition of muscles, hepatopancreas and ovary of freshwater prawn, *Macrobrachium dayanum* (Henderson). M. Phil, Thesis, Jammu University.
- Subaramanyam, C. B (1963). A note on the reproductive cycle of prawn *Peaneus indicus* of Madras Coast. *Curr Sci*, 32, 165-166.
- Subaramanyam, T. (1979). Some aspects of reproductive ecology of a male crab, *Ererita asiiatica* (M. Edwards). *J Exp Biol Eco*, 36, 259-266.

Cite This Article: Krishan Raj Kant, Chanchal Angral, Wahied Khawar Balwan (2024). Histological Study on Ovarian Development of Freshwater Prawn *Macrobrachium dayanum* (Henderson). *East African Scholars J Agri Life Sci, 7*(5), 70-75.